



## Experimental

**Samples:** Approximately 1 g of spice was weighed and extracted for 30 minutes with a quaternary solvent mixture of H<sub>2</sub>O/MeOH/ACN/THF (9:1:5:5 v/v/v/v).

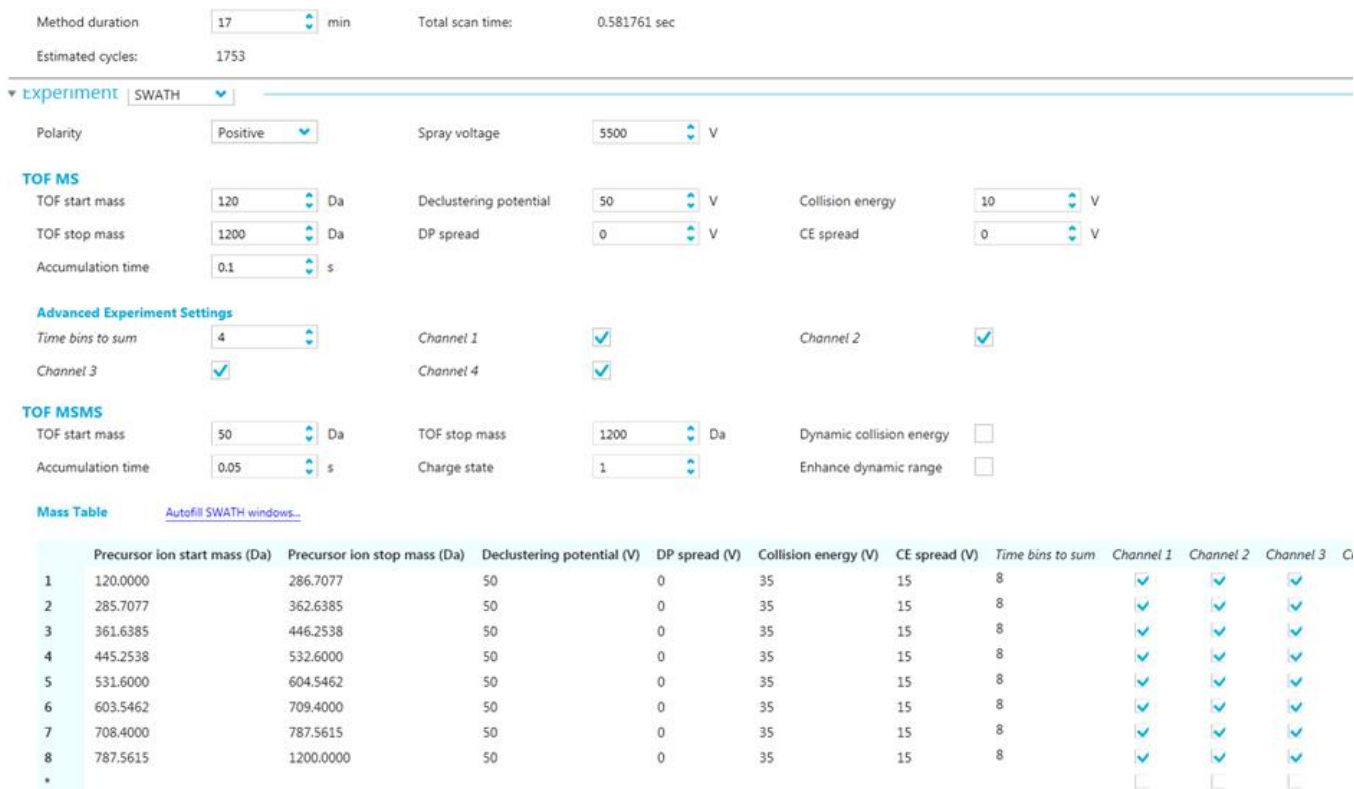
The solution was centrifuged for 5 minutes at 2500 rpm and an aliquot of the supernatant was then filtered using a 0.2 µm PTFE filter into an amber LC-vial containing three internal standards (Sudan I-d5, Sudan III-d6, Congo Red-d8).

**LC Separation:** 2 µL of the spice extracts were injected onto an ExionLC™ AD system coupled to an X500R QTOF system equipped with the TwinSprayer interface. Separation was performed using a gradient on a Waters BEH UPLC column (1.7 µm, 2.1 x 100 mm) using a mobile phase consisting of an ammonium acetate 10mM buffer (A) with methanol (B) at a flow-rate of 0.5 mL/min and column temperature of 50 °C. Gradient conditions are listed in Table 1.

**Table 1: Gradient Conditions used for the LC Separation and Subsequent Identification of the Target Dyes.**

Step	Time (min)	A (%)	B (%)
0	0.0	98	2
1	1.0	98	2
2	11.0	5	95
3	13.0	1	99
4	13.5	1	99
5	13.6	98	2
6	17.0	98	2

**SWATH Acquisition Method:** Analyses were performed using the Turbo V™ source in both negative and positive modes. The source temperature was set at 500 °C, the ion source gases 1 and 2 at 45 [AU], the curtain gas at 35 and the CAD gas at 7 [AU]. For the positive mode, the spray voltage was set at 5.5 kV and for the negative mode at -4.5 kV.



Method duration: 17 min    Total scan time: 0.581761 sec  
Estimated cycles: 1753

Experiment: SWATH

Polarity: Positive    Spray voltage: 5500 V

**TOF MS**  
TOF start mass: 120 Da    Declustering potential: 50 V    Collision energy: 10 V  
TOF stop mass: 1200 Da    DP spread: 0 V    CE spread: 0 V  
Accumulation time: 0.1 s

**Advanced Experiment Settings**  
Time bins to sum: 4    Channel 1:     Channel 2:   
Channel 3:     Channel 4:

**TOF MSMS**  
TOF start mass: 50 Da    TOF stop mass: 1200 Da    Dynamic collision energy:   
Accumulation time: 0.05 s    Charge state: 1    Enhance dynamic range:

Mass Table    Autofill SWATH windows...

	Precursor ion start mass (Da)	Precursor ion stop mass (Da)	Declustering potential (V)	DP spread (V)	Collision energy (V)	CE spread (V)	Time bins to sum	Channel 1	Channel 2	Channel 3	C
1	120.0000	286.7077	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
2	285.7077	362.6385	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
3	361.6385	446.2538	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
4	445.2538	532.6000	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
5	531.6000	604.5462	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
6	603.5462	709.4000	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
7	708.4000	787.5615	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
8	787.5615	1200.0000	50	0	35	15	8	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	
*								<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	

**Figure 2. MS Method Setup in SCIEX OS software.** The SWATH Acquisition is defined by the precursor mass windows and the MS/MS acquisition parameters.

The TOF-MS survey scan was performed from 120 to 1200 Da using these parameters for the positive mode: the declustering potential was set at 50, the accumulation time at 0.1 second and the collision energy at 10 V. For the negative mode, the declustering potential was set at -80, the accumulation time at 0.1 second and the collision energy at -10 V.

Analytes were detected by SWATH Acquisition using eight windows according to table presented in Figure 2. The following parameters were used in positive mode: the declustering potential was set at 50, the collision energy at 35 V with a spread of 15 V. For the negative mode, the declustering potential was set at -80 and the collision energy at -35 V with a spread of 15 V. The accumulation time for both modes was 0.05 seconds. The variable SWATH MS windows were optimized by evaluating the ion density over the whole chromatographic range of twelve different spices (paprika, curcuma, sweet paprika, hot chili) and spice blends (curry, satay, tandoori, garam masala, couscous ras-el-hanout, Cajun and a "seven spices" mix).

**Library:** In order to create the in-house library, and since most dyes had purity levels below 95%, they were injected using the LC method described above in IDA mode. This had the advantage of ensuring that the MS spectra were of the highest purity available. The MS/MS libraries were built at six different energy collisions: 20, 30, 40, 50 V and 35 +/-15 V and 40 +/-20 V. In the case of compounds that could be ionised in both positive and negative modes, both were added to the library.

**Data Processing:** Data was processed using the SCIEX OS software. The set-up of the peak finding criteria was done using Analytics. The criteria for the traffic lights which allows for data review and filtering can be seen in Figure 3.

## Result and Discussion

### Validation

Forty-one compounds were selected for the validation based on their color and their relevance to the study (mostly Sudan-type dyes and artificial ones). Two spices were used for validation: ground paprika and curry. Both extracts were spiked in a manner that they contained between one and thirty-two compounds. In total, forty vials were thus prepared with each analyte added randomly in twenty of them.

Configure the confidence levels for the qualitative rules, as applicable

Apply	Qualitative Rule	Acceptable Difference	Marginal Difference	Unacceptable Difference	Combined Score Weight (%)	
<input checked="" type="checkbox"/>	Mass Error (ppm)	< 5	< 10	>= 10	40	
<input checked="" type="checkbox"/>	Fragment Mass Error (ppm)	< 5	< 15	>= 15	30	
<input checked="" type="checkbox"/>	Error in Retention Time	< 0.05	< 0.1	>= 0.1	0	<input type="radio"/> Error %
<input checked="" type="checkbox"/>	% Difference Isotope Ratio	< 5	< 20	>= 20	15	<input checked="" type="radio"/> Absolute
<input checked="" type="checkbox"/>	Library Hit Score	> 70	> 50	<= 50	15	
<input type="checkbox"/>	Formula Finder Score	> 80	> 60	<= 60	20	

**Figure 3. Qualitative Rules Applied for Data Processing.** These are user-defined means of flagging results as confident matches within a set of acceptable tolerance limits for multiple parameters.

As established for screening methods, selectivity and specificity as well as the false positive and false negative rates were determined with these solutions. The vials were injected onto the LC-MS using, throughout the sequence, the integrated calibrant delivery system (CDS) with the TwinSprayer probe to maintain the mass accuracy.

After reprocessing the false positive rate was determined as 0% for all compounds whereas the false negative rate was 0% except for Amaranth (E123) and Reactive Red 195 with rates of 10% and 5%, respectively. These results highlight the excellent identification capabilities of the instrument. The mass error of the precursor ion did not exceed +/- 2 ppm for 81% of measurements (out of 494 in total) in the negative mode and 63% in the positive mode (out of 646). Only 2% of the negative mode measurements (3% in the positive mode) were comprised between +/- 5 and +/- 10 ppm and none were above +/- 10ppm.

Intra-day repeatability was assessed using both curry and paprika extracts and a representative subset of compounds. It was determined by injecting ten times the same vial in each mode successively. The parameters monitored were the retention time (RT), the raw area, the mass error and the false negative rate at the detection level. The coefficients of variation (CV) of the RT were in average below 1% for all compounds, except for Tartrazine and Acid Yellow which were nonetheless lower than 5%. In terms of false negative results, Para Red did not meet the criteria for detection (n=10) in one instance as the mass error was higher than 5 ppm at -6.4 ppm in the negative mode. Similar results were obtained with the paprika extract.

**Table 2: Intraday repeatability and summary of the results obtained in terms of raw area and number of false negatives for the curry extract.**

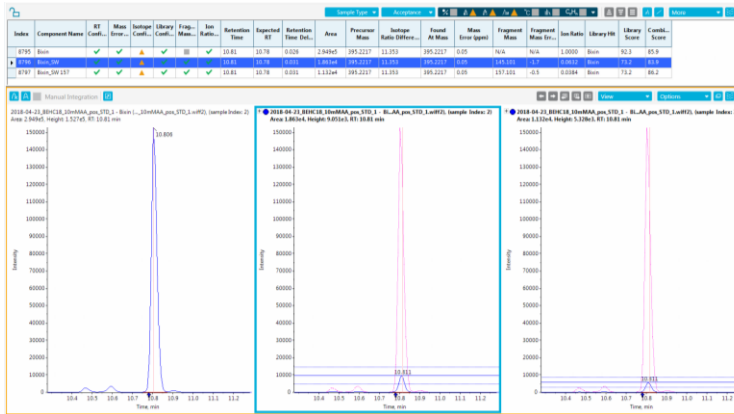
Compound	Area (average)	CV (%)	Number of False Negatives
Acid Yellow 9 (pos)	2640	21%	0
Erythrosine (neg)	9729	4%	0
Erythrosine (pos)	1004	9%	0
Para red (neg)	9248	7%	0
Para red (pos)	3120	11%	1
Ponceau 6R (neg)	34761	2%	0
Ponceau 6R (pos)	30864	4%	0
Sudan IV (pos)	18413	18%	0
Sunset Yellow (neg)	1535	7%	0
Sunset Yellow (pos)	547	10%	0
Tartrazine (neg)	1995	8%	0

## Summary

A new screening analytical method was developed to detect and identify both lipophilic Sudan-type illegal dyes and the hydrophilic artificial ones in spices. A simple solvent extraction was used and a common LC method was optimized for the analysis of 98 dyes that were added to the custom library. Forty-one colours and dyes were selected for the validation. A high degree of mass accuracy was obtained with the X500R LC-QTOF system at sufficient mass resolution regardless of the matrix type. The screening method was applied to spice samples. Sudan IV, an illegal dye, was identified with a high confidence level in a paprika sample whereas a natural food colour, bixin, was detected in a couscous spice blend. The concurrent SWATH acquisition and TOF-MS data provided excellent means of identification thanks to the accurate mass of the fragments and their ion ratios in addition to the accurate mass of the precursor ion and its' isotopic distribution. Furthermore, this type of acquisition would also allow for the retrospective analysis of suspect samples should a new “emerging” dye appear.

## Results with samples

More than 80 spice samples were purchased by local authorities from various markets and supermarkets. The traffic lights system was used to filter the data for a quick and efficient review (see Figure 3).



**Figure 4. Ion Ratios Obtained for by Bixin with Two SWATH Acquisition Fragments.**





**Figure 5: Qualitative Results for a Sample Contaminated by Sudan IV.** A) Chromatographic peak showing precursor and retention time. B) High resolution MS data including matching the observed isotope pattern to that of the target analyte. C) Matching of the MS/MS spectrum collected by SWATH acquisition to that of the target analyte in the spectral database.

## References

1. Elliot et al., Critical reviews in Food Science and Nutrition, 2017, **57** (3), 524-548
2. Colours Additives for Foods and Beverages, Ed. by M. J. Scotter, Woodhead Publishing, 2015
3. Review the toxicology of a number of dyes illegally present in food in the EU, The EFSA Journal, 2005, **263**, 1-71

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